ABSTRACT

Background: Recent studies have found some genetic variants as a risk factor for autoimmune diseases such as Multiple Sclerosis (MS). Cluster of Differentiation 226 (CD226) is one of the risk factors for MS.

Objectives: The present study aimed to evaluate the gene expression of CD226, and its protein serum level in peripheral blood samples of MS patients and healthy individuals.

Materials & Methods: A total of 30 individuals with MS and 30 healthy individuals, as controls, referred to Kashani Hospital of Isfahan, Iran. CD226 expression at the transcript level and serum protein levels were measured by quantitative real-time polymerase chain reaction and enzyme-linked immunosorbent assays, respectively. Statistical analyses were performed by Shapiro-Wilk test and nonparametric tests in SPSS.

Results: The present study showed no significant differences in the gene expression of CD226 (P=0.341). The mean serum protein level of CD226 was not different between the patients and the controls (P=0.978).

Conclusion: Overall, CD226 expression has no diagnostic usefulness in MS at either the transcript or serum level.

Keywords: Multiple Sclerosis, Demyelinating diseases, Gene expression, Cluster of Differentiation 226 (CD226) antigen
Introduction

Multiple Sclerosis (MS) is a chronic disabling inflammatory, and demyelinating disease, caused by immune responses directed against myelin proteins and progressive axonal loss in the Central Nervous System (CNS) [1]. MS is one of the most common causes of primary neurological disability in young people, and its prevalence rises with increasing age. Studies have found some genetic variants as a risk factor for MS. The present study aimed to evaluate the gene expression of CD226, and its protein serum level in peripheral blood samples of MS patients and healthy individuals. In the present study, we found no significant expression changes of CD226 in the blood samples of patients with MS. Thus, measuring the expression of CD226 gene is questionable as a biomarker for the diagnosis or improvement of MS.

Highlights

- There is no significant difference in the level of CD226 gene expression in peripheral blood sample of new cases of relapsing remitting MS patients and healthy people.
- Assessing gene expression showed no significant changes in CD226 protein expression levels.

Plain Language Summary

Multiple Sclerosis (MS) is one of the most common causes of primary neurological disability in young people, and its prevalence rises with increasing age. Studies have found some genetic variants as a risk factor for MS. The present study aimed to evaluate the gene expression of CD226, and its protein serum level in peripheral blood samples of MS patients and healthy individuals. In the present study, we found no significant expression changes of CD226 in the blood samples of patients with MS. Thus, measuring the expression of CD226 gene is questionable as a biomarker for the diagnosis or improvement of MS.
binds 2 different cell surface ligands, including poliovirus receptor (CD155) and Nectin-2 (CD112) [20].

CD112 and Necl-5 (CD155) are CAM (Costimulatory Activating Molecule) members, that form homodimers (for nectin-2) or heterodimers in their functions for cell adhesion [18]. The interaction of DNAM-1 (CD226) with its ligands is implicated in the functions of a variety of immune cells. Nectin-2 can stimulate the reaction of NK cells and cytolytic T lymphocytes through its interaction with DNAM-1 [21]. CD226 costimulatory signals potentially promote Th1 differentiation [22], but it cannot lead to differentiation of neither Th2 nor Th0 cells, enhancing IFN-γ production by naive T cells [23].

Another study reported that knockdown of CD226 on human T cells resulted in a decrease in T-bet and IFN-γ expression. However, the role of CD226 on Th2 and Th17 cells remains unknown [23]. The expression of DNAM-1 in Experimental Autoimmune Encephalomyelitis (EAE) and DNAM-1 were exhibited in the skin of patients with Systemic Sclerosis (SSc) [24]. CD226 is involved in the up-regulation of T cells. Treatment with anti-CD226 in vivo results in a significant reduction of Th1 cell expansion and in the induction of antigen presenting cells that prohibit T cell activation [25].

In total, CD226 could be an important biomarker in differentially regulating the pro-inflammatory (Th1/Th17)/anti-inflammatory (Th2) balance, indicating that the CD226 could be targeted in therapeutic approaches to autoimmune diseases like MS [17]. Therefore, the present study aimed to evaluate gene expression level of CD226 and its serum levels in patients with MS.

Materials and Methods

Study participants

A total of 30 healthy individuals and 30 new cases of Relapsing-Remitting MS (RRMS) patients diagnosed according to McDonald’s criteria were included in this study. The exclusion criteria for selecting healthy controls were as follows: Suffering from any autoimmune disease, previous organ transplantation and suffering from any inflammatory disease according to the results of erythrocyte sedimentation rate and C-reactive protein. None of the patients experienced relapse within 3 months prior to the onset of the study and were not on corticosteroid agents for at least 3 months before the onset of the research. Patients on corticosteroid therapy, and immunosuppressive drugs were excluded due to the possible effect of these agents on CD226 expression and its serum level. Table 1 presents the demographic characteristics of the participants.

Determination of CD226 mRNA expression levels

Blood samples (3 mL) were collected from all participants using an EDTA collection tube. Total RNA was extracted from the whole blood samples using a Total RNA Extraction Mini Kit (Yekta Tajhiz, Tehran, Iran). After isolation, the quality of RNA was checked by gel electrophoresis, and RNA quantity was measured using nanodrop (OD 260 nm) (Nanophotometer Pipette, Helmholtz, Nauenberg, Germany).

At the reverse transcription step, 5 ng of total RNA was used to synthesize the complementary DNA with oligo (dT) primer using the RevertAid First Strand cDNA Synthesis Kit (Thermo Scientific, USA). Quantitative real-time PCR analyses were performed using RealQ Plus 2x Master Mix Green (Ampliqon, Denmark) and Step One Plus Real-Time PCR System (Applied Biosystems).

β-Actin Gene (ACTB) was used as an endogenous control and samples were run in triplicate. Specific primers for CD226 was obtained from the report of Ye Xal. et al. and primers for ACTB as the housekeeping gene was designed by Allele ID 7.6 and BLAST (NCBI online server) [26]. Table 2 lists the sequences of primers. The relative amount of target mRNA expression was estimated by the comparative 2^ΔΔCT method which normalizes the copy number of the target mRNA to that of an endogenous reference gene (ACTB) [27].

Table 1. Demographic characteristics in the study groups

<table>
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<tr>
<th>Characteristic</th>
<th>Group</th>
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<tr>
<td></td>
<td>Case</td>
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<tr>
<td>Female/Male</td>
<td>24/6</td>
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<tr>
<td>Age, y</td>
<td>28.97±1.24</td>
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Age is represented as Mean±SD.
Determination of CD226 serum level

Blood samples (3 mL) were collected from the patients with MS and the controls, using serum-separating tubes. The blood samples were centrifuged at 1500×g for 4 min and the cell-free sera were stored at -80ºC for the Enzyme-Linked Immunosorbent Assay (ELISA). The soluble CD226 levels were measured in duplicate wells using human CD226 ELISA kit (Eastbiopharm Cat. No.: CK-E91994) according to the manufacturer’s instructions. The standard curve was drawn using the derived values based on data from the ELISA reader (Hiperion, Germany). Optical Density (OD) at 450 nm wavelength was recorded in the blood samples of both groups. The sensitivity was 2.4 ng/L.

Statistical analysis

The Shapiro-Wilk test was used to study the normality of the data. The data were then analyzed by the Mann-Whitney U test. For all evaluations, P<0.05 was considered as statistically significant. All statistical analyses were performed using SPSS.

Results

Levels of CD226 gene expression

The mean number of CD226 mRNA expressing blood was 7 in MS and was 6.76 in healthy controls. We compared CD226 gene expression between healthy controls and new cases of MS in which none of the patients of this group experienced relapse during recent 3 months and were not on corticosteroid agents since at least recent 3 months. As shown in Figure 1, even though CD226 expression increased in the patients, the difference was not statistically significant (P=0.341).

<table>
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<th>Table 2. Sequences of primers</th>
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<tr>
<td><strong>Primer Name</strong></td>
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<tr>
<td>------</td>
</tr>
<tr>
<td>CD226 F</td>
</tr>
<tr>
<td>CD226 R</td>
</tr>
<tr>
<td>ACTB F</td>
</tr>
<tr>
<td>ACTB R</td>
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<th>Table 3. Comparison of the serum levels of CD226 protein (ng/L) in both studied groups</th>
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<td><strong>Marker</strong></td>
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<td>-------------</td>
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<tr>
<td>Serum levels of CD226 protein</td>
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**CD226 serum level**

Soluble CD226 was detected in the serum of all patients with MS and healthy subjects. The mean values of CD226 serum level were 2.19 ng/mL in the case group and 2.18 ng/mL in the control group. We did not detect any significant difference of CD226 serum levels in the MS and control groups (P=0.978) (Table 3). The assessment of protein expression by ELISA confirmed the result of real-time technique. There was a correlation between the numbers of CD226 mRNA expressing in the whole blood and serum levels of soluble CD226 (P<0.05).

**Discussion**

Multiple Sclerosis is an autoimmune and neurodegenerative disease of the CNS. CD226 is involved in the upregulation of Th1 and Th17 cells. Using flow cytometry for recognizing the exact cells that express CD226 is important, because Th1 and Th17 cells play the most important role in pathogenesis of MS [28]. Here, we investigated expression of CD226 in RRMS.

Our results showed no statistically significant difference in the level of CD226 gene expression in peripheral blood sample of new cases of RRMS patients and healthy individuals. In contrast to our data, Gross et al. demonstrated that CD226 expression reduced in MS patients. They also concluded that the higher threshold for NK-cell activation is attributable to the reduced CD226 expression [29].

Another study also showed the association of non-synonymous exchange (Gly307Ser) in the gene for CD226 variant with SSc and Wegener’s Granulomatosis (WG) also demonstrated that (Single Nucleotide Polymorphism) SNPs located at CD226 gene, such as rs727088 and rs763361, can influence CD226 mRNA levels and different variant of these alleles can be protective or predispose to autoimmune diseases [30, 33]. The SNPs of each person along with the gene expression of CD226 are necessary for the exact conclusion [34].

The data regarding gene expression showed no significant changes in CD226 protein expression levels. Since the coinhibitory receptor T cell Ig and ITIM domain (TIGIT) and the costimulatory factor CD226 bind to the common ligand CD155 and the TIGIT transduce inhibitory signals and compete with CD226 for binding with CD155 on the surface of antigen presenting cells [35, 36]. It also seems necessary to demonstrate further research on the gene expression of TIGIT.

The lack of association in this study also may be due to sample size or the course of MS disease, as others considered a greater sample size in their studies. Extensive variation in the prevalence of MS in different geographical districts and races may also contributed to the difference in the result of this study compared to other studies.

**Conclusion**

We determined no significant expression change of CD226 in the blood samples of patients with MS. Therefore, measuring the expression of CD226 gene is questionable as a biomarker for the diagnosis or improvement of MS. Since CD226 targeting would exclusively target proinflammatory Th1 and Th17 cells, and these cells have the most important role in pathogenesis of MS [28], additional studies are therefore required to measure CD226 expression in certain subsets.

**Ethical Considerations**

**Compliance with ethical guidelines**

All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee. This study was approved by the Ethics Committee of Isfahan University of Medical Sciences (Grant No: 396067).

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**Conflict of interest**

The authors certify that they have no affiliation with or involvement in any organization or entity with any financial interest, or non-financial interest in the subject matter or materials dismissed in this manuscript.

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**References**


tuted by IL-2 receptor modulation. Proc Natl Acad Sci India Sect B Biol Sci. 2016; 113(21):E2973-E82. [DOI:10.1073/pnas.1524924113] [PMID] [PMCID]


